

CHROM. 7700

Note

δ -Aminolaevulinic acid synthetase

Synthesis of δ -aminolaevulinic acid pyrroles and their separation by thin-layer chromatography

M. W. ROOMI

Department of Pharmacology, Queen's University, Kingston, Ontario (Canada)

(Received April 9th, 1974)

δ -Aminolaevulinic acid (ALA) synthetase is the rate-limiting enzyme of porphyrin biosynthesis in animal, bacterial and plant cells¹⁻¹⁰. It catalyses the synthesis of δ -aminolaevulinic acid by the condensation of glycine and succinyl coenzyme A^{1,11}. The activity of this enzyme is increased in the livers of patients with acute intermittent porphyria¹², a genetic abnormality of the regulation of heme synthesis, and in the livers of animals rendered porphyric by means of variety of drugs, referred to as experimental porphyria¹³.

Granick and Urata¹⁴ induced an experimental porphyria in guinea pigs by the administration of 3,5-diethoxycarbonyl-1,4-dihydro-2,4,6-trimethylpyridine (DDC) and showed that there is a large increase in porphyrin synthesis resulting from a greater increase in the ALA-synthetase activity in liver mitochondria. Later, working with the elegant chick embryo liver cells technique, Granick¹⁵ showed that the increased activity of this enzyme, induced by DDC, results from an increased synthesis of this enzyme, rather than from activation of the inactive enzyme. ALA-synthetase activity is usually measured by converting the ALA produced by the enzyme during incubation into a pyrrole by condensation with either ethyl acetoacetate or acetylacetone¹⁶. The pyrrole formed is reacted with Ehrlich's reagent and measured. This interest in ALA-synthetase has prompted us to study ALA pyrroles. The purpose of this paper is to describe the synthesis of ALA pyrroles from δ -ALA acid hydrochloride and β -keto esters and the use of TLC to characterize these compounds. The separation by TLC of ALA pyrroles does not seem to have been studied. This study has given important information regarding the effect of various ester and alkyl substituents in the 3- and 2-positions on the mobility of the ALA pyrroles during TLC separation.

MATERIALS AND METHODS

Benzyl acetoacetate was obtained by the method described in the literature¹⁷. Methyl, ethyl and *tert.*-butyl acetoacetate and ethyl propionyl and ethyl butyryl acetate were purchased from Aldrich (Milwaukee, Wisc., U.S.A.). δ -Aminolaevulinic acid hydrochloride was synthesized according to the method of Shemin *et al.*⁵. Ultra-violet absorption spectra were determined in absolute ethanol in a Unicam SP-800

spectrophotometer. Infrared spectra were obtained in Nujol on a Perkin-Elmer Model 137E infrared spectrometer. All melting points are uncorrected.

Standard conditions for the synthesis of ALA pyrroles

A solution of β -diketone (0.01 mole) in 20 ml of phosphate buffer (0.25 M; pH 6.6) was treated with equimolar amount of δ -aminolaevulinic acid hydrochloride (0.01 mole) and refluxed for 15 min. The solution was cooled and pH was adjusted to 7. The pyrrole formed was separated by filtration, washed and crystallized from aqueous ethanol.

Thin-layer chromatography (TLC) was carried out according to Roomi^{18,19}. A suspension of 30 g of silica gel G (E. Merck, Darmstadt, G.F.R.) in 60 ml of water was spread on glass plates (20 \times 20 cm) to a thickness of 250 μ m with a Desaga applicator. The plates were dried at 105–110° for 30 min and stored in a desiccator. A 0.1% solution of the compounds in methanol was prepared and 2 μ g of each compound was spotted 2 cm from the edge of the plate. The plate was then developed with 150 ml of the solvent system diethyl ether–*n*-hexane containing 2% glacial acetic acid (1:1). Usually 45 min were required for the solvent to travel a distance of 12–15 cm. The plates were then dried, sprayed with Ehrlich reagent²⁰ and, after keeping for 5 min at 100°, the pyrroles appeared as blue spots.

RESULTS AND DISCUSSION

Fig. 1 lists the various β -diketo esters used and ALA pyrroles formed and their separation by TLC. The UV and IR spectra of these compounds are recorded in Table I. The IR spectra of all the compounds showed bands around 3300 cm^{-1} due to the N–H and at 1710 and 1660 cm^{-1} due to C=O. The UV spectra of these compounds showed a maximum around 233 nm and a minimum around 260 nm. Reaction of these pyrroles with modified Ehrlich's reagent¹⁶ gave a pink solution, which showed a maximum at 552 nm and a shoulder at 525 nm. Fig. 2 shows the absorption spectra of ALA pyrrole obtained with *tert*-butoxy ester, which has

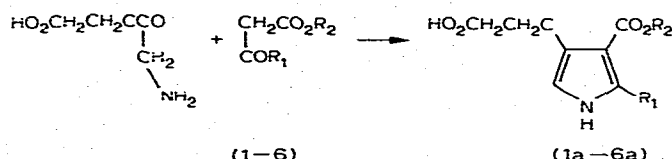


Fig. 1. Reaction scheme for synthesis of ALA pyrroles and their separation by TLC. Solvent system used: diethyl ether–*n*-hexane containing 2% glacial acetic acid (1:1).

No.	Pyrrole		$R_F \times 100$
1a	$R_1 = \text{CH}_3$	$R_2 = \text{CH}_3$	24
2a	$R_1 = \text{CH}_3$	$R_2 = \text{C}_2\text{H}_5$	29
3a	$R_1 = \text{CH}_3$	$R_2 = \text{CH}_2\text{C}_6\text{H}_5$	33
4a	$R_1 = \text{CH}_3$	$R_2 = \text{C}(\text{CH}_3)_3$	43
5a	$R_1 = \text{C}_2\text{H}_5$	$R_2 = \text{C}_2\text{H}_5$	38
6a	$R_1 = \text{C}_3\text{H}_7$	$R_2 = \text{C}_2\text{H}_5$	43

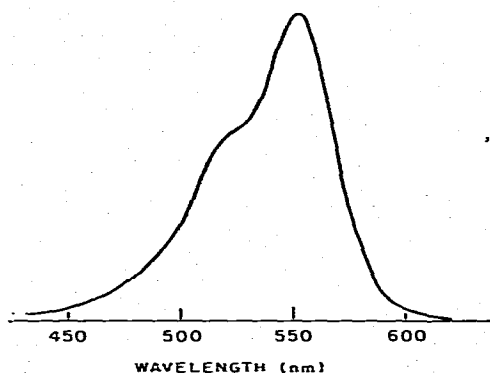


Fig. 2. Absorption curve for *tert.*-butoxy ALA pyrrole.

absorption maximum at 552 nm. The mechanism and the steps involved in the colour reaction of ALA pyrroles with Ehrlich's reagent are shown in Fig. 3.

The separation of ALA pyrroles (1a–6a) by TLC is shown in Fig. 1. 2-Methyl-3-carbethoxy-4-propionic acid pyrrole (2a) has higher R_F value than the 2-methyl-3-carbomethoxy-4-propionic acid pyrrole (1a) and are readily separated. Replacement of 2-methyl in 2a by either ethyl (5a) or propyl (6a) makes it less polar and it therefore then moves faster.

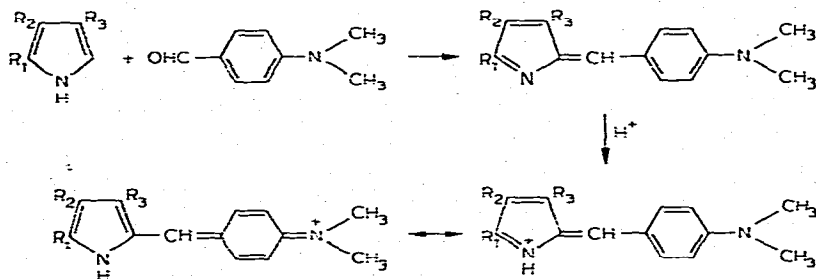


Fig. 3. Mechanism of colour reaction of ALA pyrroles with Ehrlich's reagent.

Similar substitution of 3-carbethoxy in 2a by more bulky groups, *viz.*, *tert.*-butoxy (4a) and benzyl (3a), also decreases the polarity of the compound and results in an increase R_F value. Hence the ALA pyrroles with different esters groups (methyl (1a), ethyl (2a), benzyl (3a) and *tert.*-butoxy (4a)) in the 3-position are separated from each other. Similar resolution was achieved among the ALA pyrroles having different alkyl substituents (methyl (2a), ethyl (5a) and propyl (6a)) in the 2-position. Thus all the ALA pyrroles synthesized in the present investigation can be separated from each other with the exception of 4a and 6a, which have the same R_F value.

Recently, ALA-synthetase activity has been measured in tissue obtained from liver biopsies²¹ and in cultured cells^{22,23}. The small amounts of tissue obtained by these procedures generate a small amount of ALA. The assay procedure utilizes ethyl acetoacetate to form the pyrrole prior to addition of Ehrlich's reagent. As *tert.*-buty

acetoacetate ester gives a better yield than ethyl acetoacetate when used to form the ALA pyrrole, this compound may be more desirable to use in assays of ALA-synthetase.

ACKNOWLEDGEMENTS

The author thanks Dr. G. S. Marks for his interest in this work, and the Medical Research Council, Canada, for financial support.

REFERENCES

- 1 D. Shemin and C. S. Russel, *J. Amer. Chem. Soc.*, 75 (1953) 4873.
- 2 A. Neuberger and J. J. Scott, *Nature (London)*, 172 (1953) 1093.
- 3 E. I. B. Dresel and J. E. Falk, *Nature (London)*, 172 (1953) 1185.
- 4 D. Shemin, T. Abramsky and C. S. Russel, *J. Amer. Chem. Soc.*, 76 (1954) 1024.
- 5 D. Shemin, C. S. Russel and T. Abramsky, *J. Biol. Chem.*, 215 (1955) 613.
- 6 E. I. B. Dresel and J. E. Falk, *Biochem. J.*, 63 (1956) 80.
- 7 J. E. Falk, E. I. B. Dresel, A. Benson and B. C. Knight, *Biochem. J.*, 63 (1956) 87.
- 8 S. Granick, *Science*, 120 (1954) 1105.
- 9 D. Shemin, *Methods Enzymol.*, 5 (1962) 883.
- 10 B. F. Burnham and J. Lascelles, *Biochem. J.*, 87 (1963) 462.
- 11 J. G. Wriston, L. Lack, Jr. and D. Shemin, *J. Biol. Chem.*, 215 (1955) 603.
- 12 D. P. Tschudy, M. G. Perloth, H. S. Marver, A. Collins, G. Hunter and M. Recheigl, *Proc. Nat. Acad. Sci. U.S.*, 53 (1965) 841.
- 13 R. Schmid, in J. B. Stanbury, J. B. Wyngaarden and D. S. Fredricksen (Editors), *The Metabolic Basis of Inherited Disease*, McGraw-Hill, New York, 1962, p. 939.
- 14 S. Granick and G. Urata, *J. Biol. Chem.*, 238 (1963) 821.
- 15 S. Granick, *J. Biol. Chem.*, 241 (1966) 1359.
- 16 D. Mauzerall and S. Granick, *J. Biol. Chem.*, 219 (1956) 435.
- 17 R. F. Bacon, *Amer. Chem. J.*, 33 (1905) 68.
- 18 M. W. Roomi, *J. Chromatogr.*, 65 (1972) 580.
- 19 M. W. Roomi, *J. Chromatogr.*, 70 (1972) 179.
- 20 J. E. Falk, *Porphyrins and Metalloporphyrins*, Elsevier, Amsterdam, 1964, p. 160.
- 21 R. D. Levere, E. L. Giten and S. Sassa, *J. Lab. Clin. Med.*, 75 (1970) 137.
- 22 S. Sassa and S. Granick, *Proc. Nat. Acad. Sci. U.S.*, 67 (1970) 517.
- 23 D. L. Tyrrell and G. S. Marks, *Biochem. Pharmacol.*, 21 (1972) 2077.